

Protective effects of hesperidin against genotoxicity induced by ^{99m}Tc -MIBI in human cultured lymphocyte cells[☆]

Seyed Jalal Hosseinimehr^{a,*}, Amirhossein Ahmadi^a, Davood Beiki^b,
Emran Habibi^a, Aziz Mahmoudzadeh^c

^aDepartment of Radiopharmacy, Faculty of Pharmacy and Traditional and Complementary Medicine Research Center, Mazandaran University of Medical Sciences, Sari, Iran

^bResearch Institute for Nuclear Medicine, Tehran University of Medical Sciences, Tehran, Iran

^cLaboratory of Cytogenetics, Novin Radiation Institute, Tehran, Iran

Received 9 February 2009; received in revised form 8 June 2009; accepted 14 June 2009

Abstract

Introduction: Radiopharmaceuticals have been widely used as nuclear tracers for myocardial perfusion imaging. The purpose of this study was to investigate the radioprotective effects of hesperidin as a flavonoid which protects against the genotoxic effects of ^{99m}Tc -MIBI in human cultured lymphocytes.

Methods: Whole blood samples from human volunteers were incubated with hesperidin at doses of 10, 50 and 100 μmol . After 1 h of incubation, the lymphocytes were incubated with ^{99m}Tc -MIBI (200 $\mu\text{Ci}/2\text{ ml}$) for 3 h. The lymphocyte cultures were then mitogenically stimulated to allow for evaluation of the number of micronuclei in cytokinesis-blocked binucleated cells.

Results: Incubation of lymphocytes with ^{99m}Tc -MIBI at this high dose induces additional genotoxicity and shown by increases in micronuclei frequency in human lymphocytes. Hesperidin at these doses significantly reduced the micronuclei frequency in cultured lymphocytes. The maximum protective effect and greatest decrease in micronuclei frequency occurred when cultures were incubated with a 100- μmol dose of 65% hesperidin.

Conclusion: This study has important implications for patients undergoing nuclear medicine procedures. The results indicate a protective role for hesperidin against the genetic damage and side effects induced by radiopharmaceutical administration.

© 2009 Elsevier Inc. All rights reserved.

Keywords: Radiopharmaceutical; Hesperidin; Genotoxicity; ^{99m}Tc -MIBI; Micronucleus

1. Introduction

Humans are probably exposed to ionizing irradiation through both external and internal contamination. In internal contamination, radioactive substances accumulate and cause irradiation of critical organs. Ionizing radiation passing through living tissues generates free radicals. Interactions of free radicals with DNA can induce DNA damage leading to mutagenesis and carcinogenesis [1]. One significant source

of internal irradiation stems from the use of radiopharmaceuticals in nuclear medicine procedures. Radiopharmaceuticals, which are often used for diagnostic and therapeutic purposes, contain at least one radionuclide and a non-radioactive ligand. Due to its ideal imaging energy, short physical half-life and ability to readily bind to a variety of compounds, technetium-99m (^{99m}Tc) is widely used for diagnostic imaging in nuclear medicine. Technetium forms chelates with different ligands, and the resulting complex is transferred to the target organ for image capture. ^{99m}Tc emits gamma rays (140.5 keV) as well as low-energy Auger and conversion electrons. ^{99m}Tc methoxy-isobutyl-isonitrile (^{99m}Tc -MIBI) is taken up by human peripheral blood lymphocytes, which are highly radiosensitive [2–4]. Taibi et al. [5] showed that ^{99m}Tc -MIBI induces genotoxic effects and apoptosis at an absorbed dose of 0.1 Gy (250 μCi) in human peripheral blood lymphocytes. This absorbed dose

[☆] This research was the subject of the Doctorate of Pharmacy thesis of Amirhossein Ahmadi as a student at Mazandaran University of Medical Sciences. It was supported by Mazandaran and Tehran Universities of Medical Sciences, grant no. 6240.

* Corresponding author. Tel.: +98 151 3261244; fax: +98 151 3261244.

E-mail addresses: sjhosseinim@yahoo.com,
sjhosseinim@mazums.ac.ir (S.J. Hosseinimehr).

from ^{99m}Tc -MIBI is more than that usually caused by nuclear medical diagnostic procedures [6].

Recently, we demonstrated that the flavonoid hesperidin has powerful protective effects against DNA damage induced by gamma irradiation in mice. It reduced the micronuclei frequency caused by external gamma irradiation in the mice [7]. Hesperidin has been reported to have many biologically protective properties, including anti-inflammatory, antimicrobial, anticarcinogenic, antioxidant and capillary strengthening effects [8]. Daflon, which contains hesperidin and a flavone called diosmin, is used to treat chronic venous insufficiency in Europe [9] and has been helpful in other chronic diseases. Although many studies have evaluated natural products as radioprotective agents in animals and humans, little is known about the effects of radioprotective agents against damage from radiopharmaceuticals.

The aim of this study was to determine the radioprotective effects of hesperidin against the genotoxicity induced by ^{99m}Tc -MIBI in human peripheral blood lymphocyte cells in vitro.

2. Material and methods

2.1. Chemicals and preparation of ^{99m}Tc -MIBI

All chemicals were obtained from Merck and Sigma, and were used without further purification. Hesperidin was prepared in sterile water and DMSO (0.25%). ^{99m}Tc , in the form of sodium pertechnetate ($\text{Na}^{+99m}\text{TcO}_4^-$), was eluted from an in-house $^{99}\text{Mo}/^{99m}\text{Tc}$ generator system with normal saline solution. A commercial sestamibi kit (AEOI, Tehran, Iran) was used, and the labeling and quality control procedures were performed according to the manufacturer's instructions.

2.2. Irradiation protocol

The study protocol was approved by the ethics committee of the university. After obtaining written informed consent, 12-ml blood samples were collected in heparinized tubes from three healthy, nonsmoking male volunteers, aged 25–35 years. The blood was divided into six 2-ml tubes, one for each of the six study groups: control; ^{99m}Tc -MIBI only; hesperidin at doses of 10, 50 and 100 μmol (final concentrations) with ^{99m}Tc -MIBI; and hesperidin only. First, blood samples were incubated with hesperidin for

1 h. Then, 200 μCi of ^{99m}Tc -MIBI was added to the blood samples and incubated for 3 h. After incubation, RPMI 1640 medium was added to each tube and the cultures were centrifuged at 1200 rpm for 8 min. To separate ^{99m}Tc -MIBI from the whole blood, the upper (less dense) solution was removed and blood was transferred for micronucleus assay.

2.3. Micronuclei assay

One-half milliliter of each sample (control and irradiated groups) was added to 4.5 ml of RPMI 1640 culture medium (Gibco, USA), which contained 20% fetal calf serum, 0.1 ml/5 ml phytohemagglutinin (Gibco), antibiotics (penicillin 100 IU/ml, streptomycin 100 $\mu\text{g}/\text{ml}$) and 2 mM glutamine (Sigma, USA) at final concentration. All cultures were set up in duplicate and incubated at $37\pm 1^\circ\text{C}$ in a humidified atmosphere of 5% $\text{CO}_2/95\%$ air. Cytochalasin B (Fluka, final concentration: 30 $\mu\text{g}/5$ ml) was added after 44 h of culture incubation. At the end of 72 h of incubation, the cells were collected by centrifugation and resuspended in 0.075 M cold potassium chloride for 8 min at 1000 rpm. They were then immediately treated with a fixative solution three times (methanol/acetic acid, 6:1). Fixed cells were dropped onto clean microscopic slides, air-dried and stained with Giemsa solution. All slides were coded by an individual other than the scorer and evaluated at $40\times$ magnification for the micronuclei frequency in cytokinesis-blocked binucleated cells with well-preserved cytoplasm. To be scored as micronuclei, candidates had to have a diameter of between 1/16 and 1/3 of the main nuclei, be nonrefractile and not be linked to or overlap with the main nuclei [10]. At each blood collection time, 1000 binucleated cells from duplicate irradiated and control cultures from each volunteer were examined to record the frequency of micronuclei.

2.4. Statistical analysis

At each blood collection, the prevalence of micronuclei was recorded for each volunteer. The data were analyzed using ANOVA with Tukey's HSD post hoc test.

3. Results

The percentage of micronuclei in the lymphocytes of volunteers treated with 200 μCi of ^{99m}Tc -MIBI was

Table 1

The percentages of micronuclei induced in vitro by ^{99m}Tc -MIBI (TC) on cultured blood lymphocytes from human volunteers and protective effects of hesperidin (HES)

Volunteer group	Binucleate cells with micronuclei (%)*					
	Control	TC	TC+HES10	TC+HES50	TC+HES100	HES100
Volunteer 1	0.7	3.8	3.2	1.3	1.3	0.5
Volunteer 2	0.7	3.3	3.8	1.5	1.1	1.0
Volunteer 3	1	3.9	4.0	2.0	1.5	1.4
Mean \pm S.D.	0.8 \pm 0.17	3.67 \pm 0.32	3.67 \pm 0.42	1.6 \pm 0.36	1.3 \pm 0.2	0.97 \pm 0.42

* One thousand BN cells were examined in each culture.

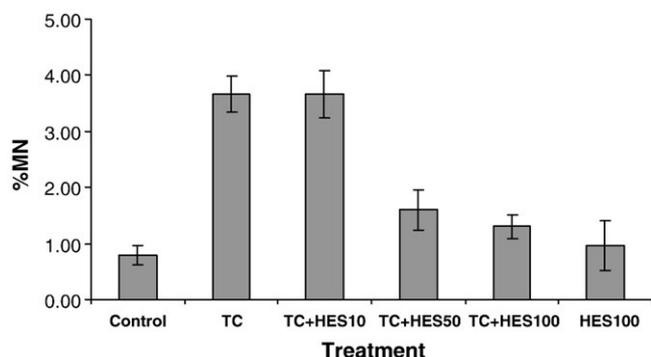


Fig. 1. In vitro protection by hesperidin (HES) at different concentrations (10, 50 and 100 μmol) against radiation-induced genetic damage induced by $^{99\text{m}}\text{Tc}$ -MIBI (TC) in cultured whole blood lymphocyte. The data represent the average \pm S.D. of three human volunteers. $P<0.0001$: Control sample compared with similarly irradiated lymphocytes from the blood sample treated with TC. $P<0.001$: TC sample compared to TC-HES50 and TC-HES100 samples. $P<0.001$: TC-HES10 sample compared to TC-HES50 and TC-HES100 samples. $P>0.05$: TC-HES50 sample compared to TC-HES100 sample.

3.67 \pm 0.32%, while the percentage in nontreated control lymphocytes was 0.80 \pm 0.17% (Table 1). The frequency of micronuclei (an indication of the genotoxic effects of internal irradiation) after preincubation with hesperidin at doses of 10, 50 and 100 μmol was 3.67 \pm 0.42, 1.6 \pm 0.36 and 1.3 \pm 0.2, respectively (Fig. 1). The data demonstrate that the frequencies of micronuclei found in the hesperidin-treated samples were significantly lower than in the samples cultured with $^{99\text{m}}\text{Tc}$ -MIBI only. Whole blood samples incubated with 50 and 100 μmol of hesperidin, and then exposed in vitro to $^{99\text{m}}\text{Tc}$ -MIBI radiation, exhibited a significant decrease in micronuclei frequency compared to those samples incubated with 10 μmol of hesperidin only ($P<0.0001$). Total micronuclei values were 56% and 65% less in the samples treated with hesperidin at concentrations of 50 and 100 μmol , respectively, than in controls (Table 1). Hesperidin did not show any protective effect at a concentration of 10 μmol . Hesperidin alone did not cause genotoxicity in cultured lymphocytes at concentrations of 100 μmol .

A typical depiction of binucleated cells with micronuclei is shown in Fig. 2.

4. Discussion

In this study, we show that hesperidin significantly protects against genotoxicity induced by the radiotracer $^{99\text{m}}\text{Tc}$ -MIBI in lymphocytes. In vitro incubation of human blood with the natural compound hesperidin reduced the frequency of micronuclei induced by internal irradiation by the radiopharmaceutical $^{99\text{m}}\text{Tc}$ -MIBI. Measures of genotoxicity have been used to estimate the risk of damage induced by internal irradiation from the radiopharmaceutical [5].

Genotoxic agents can cause cancer, hereditary disorders and abnormalities in developing embryos [11].

In this study, we have observed genotoxic effects induced by $^{99\text{m}}\text{Tc}$ -MIBI as shown by increases in lymphocytes at a dose of 200 μCi . Taibi et al. [5] studied the effect of increasing $^{99\text{m}}\text{Tc}$ -MIBI activity in vitro (corresponding to absorbed doses ranging from 1 μGy to 1 Gy) on healthy human lymphocytes. The micronuclei frequency was similar in control cultures and lymphocyte cultures exposed to doses of 10 μGy , 100 μGy and 1 cGy. A significantly higher frequency of micronuclei was observed after exposure to a dose of 10 cGy (corresponding to 250 μCi of $^{99\text{m}}\text{Tc}$ -MIBI) [5]. Although we observed an increase in the micronuclei frequency in lymphocytes treated with $^{99\text{m}}\text{Tc}$ -MIBI, this dose of radiotracer was much higher than the usual dose used for diagnosis of patients in the clinical setting. Bolus injections of 700 MBq (19 mCi) of $^{99\text{m}}\text{Tc}$ -MIBI exhibited peak activity in the blood after 30 s with about 0.00040 fraction of injected dose absorbed per milliliter [6]. Whole blood activity was about 7.6 $\mu\text{Ci}/\text{ml}$. $^{99\text{m}}\text{Tc}$ -MIBI was rapidly cleared from the blood with a total clearance rate of more than 4 L/min. $^{99\text{m}}\text{Tc}$ -MIBI reached its minimum blood concentration of about 0.00001 fraction of radioactive dose 5 min after injection [6].

A recent study reported that the micronuclei frequency does not significantly increase in patients undergoing perfusion imaging using $^{99\text{m}}\text{Tc}$ -MIBI injection as compared to controls [12]. In that study, patients received a high dose of 40 mCi $^{99\text{m}}\text{Tc}$ -MIBI for myocardial diagnosis and there was no genotoxicity observed in the cultured lymphocytes 48 h after [12]. In the current study, we used a high blood concentration of $^{99\text{m}}\text{Tc}$ -MIBI (200 $\mu\text{Ci}/\text{ml}$ of whole blood) for a model of internal irradiation similar to that in the Taibi et al. [5] study. $^{99\text{m}}\text{Tc}$ -MIBI was chosen for several reasons. First, over 80% of radiopharmaceuticals currently used in diagnostic nuclear medicine are labeled with $^{99\text{m}}\text{Tc}$, which has optimal nuclear properties for nuclear medicine imaging [3]. Decaying technetium-99m not only emits gamma rays, but also emits Auger electrons that cause

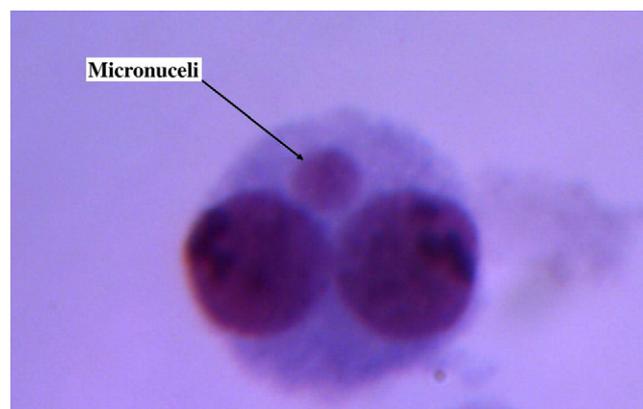


Fig. 2. A typical binucleated lymphocyte with micronuclei in our study.

subcellular side effects due to the extremely short range of action in the cell [2]. Second, myocardial perfusion scintigraphy is a routine nuclear diagnostic test for cardiac disease. In patients undergoing bone scintigraphy, additional induction of chromosomal aberrations by ^{99m}Tc -hexamethylene diphosphonate (^{99m}Tc -HDP) was not observed. ^{99m}Tc -HDP has also been reported to not cause any genotoxic effects after intravenous administration at doses of 20–25 mCi [13].

The genotoxic effects induced by radiopharmaceuticals were evaluated by incubating human lymphocytes with ^{99m}Tc during ^{99m}Tc -labeling of lymphocytes. In a study aimed at determining whether ^{99m}Tc induced genotoxicity after labeling of lymphocytes, researchers observed that increasing dose of ^{99m}Tc resulted in proportional increases in micronuclei in lymphocytes at doses of 0.236, 0.435 and 0.580 mCi per 10^7 of isolated lymphocyte cells [4]. The 0.580-mCi dose of ^{99m}Tc corresponded to 1 Gy of 250-keV X-ray emission; the frequency of micronuclei was 5.5% and 0.68% after 72 h of incubation of lymphocytes with ^{99m}Tc at a dose of 0.580 mCi per 10^7 for the samples and controls, respectively [4]. Patients exposed to clinically used doses of technetium-99m did not show any chromosomal aberrations or mutations in peripheral blood lymphocytes [14]. Internal radiation from different radionuclides, as well as external radiation of various types, can induce genotoxicity, which increases the risk of cancer. Dose–response models in animals looking at the effects of high doses may be used to assess the risks associated with ionizing radiation emitted by radiolabeled compounds in humans [11].

Hesperidin has a molecular weight of 610.5 and is found mainly in the citrus family. It has been reported to have several beneficial health effects, including chemoprotection against carcinogenesis in the colon [15,16] and reduction of oxidative stress in the rat liver and kidney [17]. The vasoprotective and venotonic effects of hesperidin make it a potential therapeutic agent for patients. Daflon (composed of hesperidin plus diosmin) is used to treat chronic venous insufficiency in Europe [9]. Ionizing radiation produces free radical damage in DNA and induces genotoxic effects and apoptosis. Free radical scavenging is apparently responsible for the inhibitory effects of flavonoids, such as rutin, morin, quercetin and genestin, on the clastogenic activity induced by gamma radiation in mice [18].

The molecular mechanisms underlying the radioprotective effects of hesperidin are not clear. Hesperidin has been shown to have antioxidant activity against the cellular oxidative stress associated with neurodegenerative diseases. This flavonoid also attenuated decreases of glutathione peroxidase and glutathione reductase activity and decreased DNA damage in H_2O_2 -induced PC12 cells [19], and also inhibited low-density lipoprotein oxidation [20]. Oral administration of hesperidin has protective effects against gamma radiation-induced hepatocellular damage and oxidative stress in rats [21]. The pharmacokinetic parameters of hesperidin were measured for total hesperidin in healthy

humans after the consumption of orange juice [22]. In that protocol, administration of orange juice corresponded to 192 mg hesperidin, with a peak plasma concentration (C_{max}) of 1.05 mmol/L of hesperitin (aglycone of hesperidin). In our study, the concentration of hesperidin (molecular weight, 610.5) was 6.1, 30.5 and 61.05 $\mu\text{g}/\text{ml}$ of incubated whole blood for 10, 50 and 100 μmol , respectively. In this study, we showed that hesperidin protects human lymphocytes against genotoxicity induced by internal irradiation. For all three volunteers, the ^{99m}Tc -MIBI-treated lymphocytes incubated with hesperidin showed a reduction in micronuclei frequency as compared to ^{99m}Tc -MIBI-treated samples without hesperidin. This finding suggests that hesperidin acts effectively as a free radical scavenger as such radioprotection is concentration dependent. Hesperidin treatment at 100 μmol provides maximum leukocyte protection.

Since hesperidin has been used extensively as a pleiotropic drug, and with regards to potential radioprotective effect, it may be a useful therapeutic candidate for patients undergoing nuclear medicine procedures.

Acknowledgment

We would like to thank Dr. Omid Beiki for his technical assistance.

References

- [1] Hosseinimehr SJ. Trends in the development of radioprotective agents. *Drug Discov Today* 2007;12:794–805.
- [2] Pomplun E, Terrissol M, Kümmerle E. Estimation of a radiation weighting factor for ^{99m}Tc . *Radiat Prot Dosim* 2006;122:80–1.
- [3] Schwochau K. Technetium radiopharmaceuticals—fundamentals, synthesis, structure and development. *Angew Chem Ed Engl* 1994; 33:2258–67.
- [4] Merz T, Tatum J, Hirsch J. Technetium-99m-labeled lymphocytes: a radiotoxicity study. *J Nucl Med* 1986;27:105–10.
- [5] Taibi N, Aka P, Krisch-Volders M, Bourgeois P, Fruhling J, Szpircer C. Radiobiological effect of ^{99m}Tc -MIBI in human peripheral blood lymphocytes: ex vivo study using micronucleus/FISH assay. *Cancer Lett* 2006;233:68–78.
- [6] Sand NP, Julsgaard P, Rasmussen K, Flo C, Thuesen L, Bagger JP, et al. Arterial concentration of ^{99m}Tc -sestamibi at rest, during peak exercise and after dipyridamole infusion. *Clin Physiol Funct Imaging* 2004;24:394–7.
- [7] Hosseinimehr SJ, Nemati A. Radioprotective effects of hesperidin against gamma irradiation in mouse bone marrow cells. *Br J Radiol* 2006;79:415–8.
- [8] Garg A, Garg S, Zaneveled JD, Singla AK. Chemistry and pharmacology of the citrus bioflavonoid hesperidin. *Phytotherapy Res* 2001;15:655–69.
- [9] Yang M, Tanaka T, Hirose Y, Deguchi T, Mori H, Kawada Y. Chemopreventive effects of diosmin and hesperidin on *N*-butyl-*N*-(4-hydroxybutyl) nitrosamine-induced urinary-bladder carcinogenesis in male ICR mice. *Int J Cancer* 1997;73:719–24.
- [10] Fenech M. The in vitro micronucleus technique. *Mutat Res* 2000;455: 81–95.
- [11] Lundqvist H, Antoni G, Långström B. Genotoxic hazard of radiopharmaceuticals in humans: chemical and radiation aspects coupled to microdosing. *Eur J Clin Pharmacol* 2007;63:641–5.

- [12] Mogharrabi M, Houman A, Mosaffa N, Tabeie F, Valaei N, Shafiee B, et al. Investigating the effect of high-dose ^{99m}Tc -MIBI on chromosomal damage induction among patients undergoing myocardial perfusion scan. *Pajoohandeh* 2008;9-15:13 [Persian].
- [13] Guiraud-Vitoux F, Jacquet N, Petiet A, Roy L, Voisin P, Colas-Linhart N. Induction of unstable and stable chromosomal aberrations by ^{99m}Tc : in vitro and in vivo studies. *Nucl Med Commun* 2005;26:913–8.
- [14] Kelsey KT, Donohoe KJ, Memisoglu A, Baxter B, Caggana M, Liber HL. In vivo exposure of human lymphocytes to technetium-99m in nuclear medicine patients does not induce detectable genetic effects. *Mutat Res* 1991;264:213–8.
- [15] Tanaka T, Kohno H, Murakami M, Shimada R, Kagami S, Sumida T, et al. Suppression of azoxymethane-induced colon carcinogenesis in male F344 rats by mandarin juice in β -cryptoxanthin and hesperidin. *Int J Cancer* 2000;88:146–50.
- [16] Tanaka T, Makita H, Kawabata K, et al. Chemoprevention of azoxymethane-induced rat colon carcinogenesis by naturally occurring flavonoids, diosmin and hesperidin. *Carcinogenesis* 1997;18: 957–65.
- [17] Tirkey N, Pilkhwai S, Kuhad A, Chopra K. Hesperidin, a citrus bioflavonoid, decreases the oxidative stress produced by carbon tetrachloride in rat liver and kidney. *BMC Pharmacol* 2005;5:1–8.
- [18] Hwang SI, Yen Gc. Neuroprotective effects of the citrus flavanones against H_2O_2 -induced cytotoxicity in PC12 cells. *J Agri Food Chem* 2008;13:859–64.
- [19] Shimoi K, Masuda S, Furugori M, Esaki S, Kinae N. Radioprotective effect on antioxidative flavonoids in γ -ray irradiated mice. *Carcinogenesis* 1994;15:2669–72.
- [20] Liu R, Meng F, Liu Y, Bai H, Liu BW. Inhibitory effect of isorhamnetin and hesperidin on LDL oxidation induced by Cu^{2+} . *Zhong Yao Cai* 2007;30:677–81.
- [21] Pradeep K, Park SH, Ko KC. Hesperidin a flavanoglycone protects against γ -irradiation induced hepatocellular damage and oxidative stress in Sprague-Dawley rats. *Eur J Pharmacol* 2008;587:273–80.
- [22] Nielsen IL, Chee WSS, Poulsen L, Offord-Cavi E, Rasmussen SE, Frederiksen H. Bioavailability is improved by enzymatic modification of the citrus flavonoid hesperidin in humans: a randomized, double-blind, crossover trial. *J Nutr* 2006;136:404–8.